# Determination of Process-related Genotoxic Impurities of Pentoxifylline by GC-HS Method

Sunil B. Lakhmapure<sup>1</sup>, Sandip A. Telavane<sup>1</sup>, Seema Kothri<sup>1</sup>, Sushil Kumar Singh<sup>2</sup>, Manohar V. Lokhande <sup>1, 2\*</sup>

<sup>1</sup> Department of Chemistry, PAHER University, Udaipur-303003, Rajasthan, India <sup>2</sup>Department of Chemistry, Sathaye College, Mumbai-400057, Maharashtra, India

## Abstract

The synthesis of pentoxifylline contained three different kinds of genotoxic contaminants. While 6-chloro-2-hexanone is possibly genotoxic, 3,7-dimethyl-1-(5-oxohexyl)-2,3,6,7-tetrahydro-1H-purine-2,6-dione is not; structural alert for genotoxic mutagenicity and carcinogenicity is present. According to the structural alert and QSTR model consensus, 1-bromo-3-chloropropane and 3-chloro-1-propanol are both genotoxic. For most volatile and semi-volatile analytes, gas chromatography (GC) with direct liquid injection and a flame ionization detector (FID) is the appropriate method. The suggested approach was shown to be very effective, precise, linear, accurate, and specific in monitoring three genotoxic contaminants. Pharmaceutical examination of 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol. GC techniques were used to analyze these contaminants. The method is particularly sensitive to quantifying 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol contaminants at 0.3 ppm and 0.9 ppm, respectively, according to the results of the LOQ and LOD methods. The goal of this work is to determine threshold amounts and appropriate methodologies for quantifying the three genotoxic contaminants that may be present during pentoxifylline production.

**Keywords:** Genotoxic impurities, gas chromatography (GC, flame ionization detector (FID), chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol

## Introduction

Pentoxifylline, sometimes referred to as oxpentifylline, is a xanthine derivative that is prescribed to patients with peripheral artery disease to relieve their muscle pain. <sup>[1]</sup> It is marketed globally under numerous brand names and is generic. <sup>[2]</sup> It is primarily used in medicine to treat intermittent claudication, a type of muscle discomfort brought on by peripheral artery disorders, which causes pain, cramps, numbness, or weakness in the arms or legs. SIGN also recommends the use of pentoxifylline as a licensed supplement to compression bandaging for the treatment of persistent venous leg ulcers. <sup>[3]</sup> as it has been demonstrated to increase healing rates. <sup>[4]</sup> Additionally, it has been demonstrated that pentoxifylline helps with alcoholic hepatitis, with some studies demonstrating a reduction in the risk of hepatorenal syndrome.

*Mechanism of action*: Pentoxifylline, one of the other methylated xanthine derivatives, is a competitive nonselective phosphodiesterase inhibitor <sup>[5]</sup> that also increases intracellular cAMP, activates PKA, suppresses leukotriene production and TNF <sup>[6-7]</sup>, lowers innate immunity and inflammation. <sup>[8]</sup> Pentoxifylline also lessens blood viscosity, enhances red blood cell deformability (a hemorheological action), and lowers the risk of platelet aggregation and thrombus formation. <sup>[9]</sup> Additionally, pentoxifylline inhibits adenosine 2 receptors.<sup>[10]</sup> Pentoxifylline may be able to reduce the levels of several biomarkers in non-alcoholic steatohepatitis, according to some research <sup>[11]</sup>, but not enough to say if this usage of the medication is safe and effective.<sup>[12]</sup> Research on the use of pentoxifylline for hearing loss and erectile dysfunction in animals has been done <sup>[13]</sup>. Studies on humans have been done for Peronei's disease.<sup>[14]</sup>

Pentoxifylline is preventive against osteoradionecrosis and repairs refractory osteoradionecrosis of the jaw when combined with tocopherol and clodronate.<sup>[15]</sup> The following was determined by a Cochrane systematic review on the use of pentoxifylline for intermittent claudication. The majority of the included studies had poor quality, and there was a lot of variation in the reported results regarding trial length<sup>[16]</sup>, pentoxifylline dosages, and participant walking lengths at the beginning of the trials. The majority of the included studies lacked sufficient information to allow for the assessment of selective reporting, did not report on the blinding of outcome assessors, and did not discuss randomization procedures or how treatment allocation was hidden. Given all these factors, the role of pentoxifylline in intermittent claudication remains uncertain, although participants generally tolerated this medication well.



#### Manufacturing Process of Pentoxifylline: (Reaction Scheme)



The literature review verified that, as of now, no techniques have been documented for identifying the genotoxic impurities of pentoxifylline, which include 1-bromo-3-chloropropane <sup>[17]</sup> (CAS no: 109-70-6) 3-chloro-2-hexanone <sup>[18]</sup> (CAS number: 627-30-5), and 6-chloro-2-hexanone <sup>[19]</sup> (CAS no: 10226-30-9). The development of FID-GC techniques for the identification of pentoxifylline and its contaminants is emphasized throughout the dissertation. Based on the genotoxic prediction report for Pentoxifylline's process impurities structures, which indicates that these impurities are genotoxic, the author felt it was necessary to identify these impurities using an appropriate chromatographic technique. As a result, this job was finished in two sections.

#### Genotoxicity prediction methods and confirmation

**Objective:** To predict the Genotoxic potential of given compounds.

*Method*: Genotoxicity prediction is a consensus inference derived from three different methodologies.

a) Decision tree-based Alerts: It uses the fragment rule base which is validated by the results of the Joint Research Centre's European Chemicals Bureau hazard estimation based on the Benigni/Bossa<sup>[20]</sup> rule base for genotoxic carcinogenicity<sup>[21-22]</sup> and mutagenicity.

**b)** Toxicophores significance by ANOVA: This methodology makes use of a database of substances from Gold <sup>[23]</sup> and Zeiger that have TD50 values given for species of rats and mice.<sup>[24]</sup> ANOVA analysis is performed using carefully selected toxicophoric data from the

literature, and the related F-ratio and probability are computed. The fragment's relevance about its contribution to genotoxicity is stated if it is found in any of the test chemicals. Estimating the degree of a compound's toxicity is made easier by the relevance of its hazardous fragments.

**c) Predicting genotoxicity using SAR / QSAR models:** The nearest neighbour method is used to create strong and trustworthy QSTR models for genotoxicity, mutagenicity <sup>[25–27]</sup>, and carcinogenicity.<sup>[28]</sup> The classification model has been verified by OECD <sup>[29]</sup> Guidance and REACH rules. <sup>[30]</sup>

#### **Materials And Methods**

a) Materials and Reagents: Methylene Chloride (GC Grade) was obtained from Merck Specialty Chemicals, Mumbai, India. Reference materials 6-chloro-2-hexanone and 3-chloro-1-propanol were obtained from Tokyo Chemical Industry, Mumbai, India and 1-bromo-3chloropropane was obtained from Alfa-Acer and Pentoxifylline was obtained from Supriya Lifescience Ltd, Mumbai.

#### Fig.1: Structure of Pentoxifylline





**b) Instrumentation:** The gas chromatographic analysis was carried out using GC 7890B equipped with a flame ionization detector (FID) and 7683B auto-injector (Agilent Technologies, USA). The chromatographic data was recorded using Agilent data acquisition software.

#### c) Preparation of Solutions:

*Blank Solution:* Methylene chloride was used for the preparation of sample and standard solutions.

*Standard Solution:* Each of 10 mg of 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol diluted with 100 ml of methylene chloride into a 100mL volumetric flask.

Stock Solution: 1.0 mL of the above solution was transferred into a 100 mL volumetric flask and diluted up to the mark with methylene chloride. This corresponds to 1 ppm of each impurity w.r.t. the test concentration.

*Pentoxifylline sample solution:* 1.0 g of Pentoxifylline sample was accurately weighed and taken into a 100 mL volumetric flask and diluted with 100 ml of methylene chloride.

## **RESULTS AND DISCUSSIONS:**

**METHOD DEVELOPMENT:** Since gas chromatography with direct liquid injection and a flame ionization detector (FID) works well with the majority of volatile and semi-volatile analytes, the author used it in their most recent work. The major difficulty was getting gas chromatography to produce the appropriate detection and quantification limit. Choosing the right solvent for pentoxifylline's solubility has turned into a problem in achieving the necessary concentration for a better FID signal. As a result, we tested several GC columns and dilution solvents in the current study to meet the goals and demands of the industry.

a) Method Development Experiment-1

Blank Solution: DMSO was used for the preparation of sample and standard solutions.

*Standard Solution:* Each of 10 mg of 6-chloro-2-hexanone, 1-bromo-3-chloropropane and 3-chloro-1-propanol diluted with 100 ml of DMSO into a 100mL volumetric flask.

Chromatographic conditions:

Column: DB-17 (Agilent), 30 m x 0.32 mm ID x 0.5µm capillary column or equivalent. (50% phenyl – methyl polysiloxane); Injection mode: Split; Flow control mode: Constant flow; Column flow: 2 ml/min; Linear Velocity: 37 cm/sec; Purge flow: 3.0 ml/min; Split ratio: 3.0; Carrier gas: Nitrogen; Column Temperature: 100°C (hold 2 min.) to 230 °C @ 30 °C (hold 5 min.) Injector Temperature: 250 °C; Detector Temperature: 260 °C Injection Volume: 1µl; Detector: FID.

*Observation:* The two genotoxic impurities were only detected and also resolution was not getting proper at the threshold level i.e. 1ppm with the above experimental conditions.

#### b) Method Development Experiment-2

*Blank Solution:* Methylene chloride was used for the preparation of sample and standard solutions.

*Standard Solution:* Each of 10 mg of 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol diluted with 100 ml of Methylene chloride into a 100mL volumetric flask.

*Observation:* The three genotoxic impurities were detected and also resolution was not getting proper at the threshold level i.e. 1ppm with the above experimental conditions.

### c) Final Chromatographic Conditions

The main target of this study was to develop a GC method to get better resolution and good response between Pentoxifylline and its three genotoxic impurities at 1 ppm level. Better results were obtained by using a DB-17 ( $30m \times 0.32$  mm internal diameter) column with a film thickness of 0.5µm and with an oven temperature program 100°C (hold 2 min.) to 230 °C @ 30

°C (hold 5 min.). Finally, the experiment was completed with the following chromatographic conditions.

*Instrument:* GC 7890B equipped with flame ionization detector (FID) and 7683B auto-injector (Agilent Technologies, USA). Carrier gas: Nitrogen Column flow

Column: DB-17 (Agilent), 30 m x 0.32 mm ID x 0.5µm capillary column or equivalent. (50% phenyl – methyl polysiloxane) ; Injection mode: Split; Flow control mode: Constant flow; Column flow: 2 ml/min; Linear Velocity: 37 cm/sec; Purge flow: 3.0 ml/min; Split ratio: 3.0; Carrier gas: Nitrogen; Column Temperature: 100°C (hold 2 min.) to 230 °C @ 30 °C (hold 5 min.) ; Injector Temperature: 250 °C; Detector Temperature: 260 °C; Injection Volume: 1µl; Detector: FID.

<u>Observation</u>: It is observed that the two genotoxic impurities were well resolved, with good response and better peak shapes at the threshold level i.e. 1ppm. The advanced developed method was validated as per ICH guidelines <sup>[32-33]</sup>. The validation parameters included selectively/ specificity, the limit of detection, the limit of quantification, accuracy, precision, linearity, ruggedness, and robustness.

*Specificity:* The capacity to evaluate the analyte without a doubt in the presence of any other traces of components that could be anticipated to be present is known as specificity/selectivity.<sup>[34]</sup> To achieve this, genotoxic contaminants were added to the Pentoxifylline sample (1.0 $\mu$ g/mL blend with Pentoxifylline test concentration of 10 mg/ml). Every impurity was well separated from the others and provided superior separation.

Fig. 2: Chromatogram for Genotoxic impurities spiked with Pentoxifylline





Fig. 3: Chromatogram for Genotoxic impurities spiked with Pentoxifylline

*THE DETECTION LIMIT (LOD) AND QUANTITATION LIMIT (LOQ)*: To determine DL and QL values, the concentration of impurities <sup>[35]</sup> was reduced sequentially such that they yielded S/N of about 3 and 10 respectively thus preparing a series of solutions of 0.1, 0.2, 0.3, 0.4 and 0.5 ppm with mixed concentrations of 6-chloro-2-hexanone, 1-bromo-3-chloropropane and 3-chloro-1-propanol and injected into the GC as described in methodology. The method was carried out by performing six replications of QL concentration and %RSD was calculated. The results indicate that the method is very sensitive to quantifying 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol impurities at 0.3 ppm and 0.9 ppm respectively.



Fig. 4: Chromatogram of LOD for Genotoxic impurities



## Fig. 5: Chromatogram of LOQ for Genotoxic impurities

Table 1: Precision of (LOD) for Pentoxifylline genotoxic impurities

	Concentration		Response (Area)			
Solution No	ppm	%	3-chloro-1-	1-bromo-3-	6-chloro-2-	
			propanol	chloropropane	hexanone	
LOD Sol-1			30173	31945	45895	
LOD Sol-2			30908	29030	49625	
LOD Sol-3	0.3	0.0003	30283	30320	47731	
LOD Sol-4			30871	30285	48970	
LOD Sol-5			34472	31738	44014	
LOD Sol-6			33917	28728	45472	
		Average	91774	87213	31771	
		Std. Dev.	3295.88	4224.79	1909.10	
		RSD	3.59	4.84	6.01	

 Table 2: Precision of LOQ for Pentoxifylline genotoxic impurities

	Concer	ntration	Response (Area)			
Solution No	ppm	%	3-chloro-1-	1-bromo-3-	6-chloro-2-	
			propanol	chloropropane	hexanone	
LOQ Sol-1			91229	84167	135529	
LOQ Sol-2			91467	89282	134226	
LOQ Sol-3	0.9	0.0009	91778	81423	137874	
LOQ Sol-4			97498	87141	135351	
LOQ Sol-5			87183	87569	133819	
LOQ Sol-6			91489	93696	132977	
		Average	91774	87213	134963	
		Std. Dev.	3295.88	4224.79	1717.03	
		RSD	3.59	4.84	1.27	

	Response (Area)				
Sr. No.	3-chloro-1-propanol	1-bromo-3-chloropropane	6-chloro-2-hexanone		
Inj. 1	99460	85238	141441		
Inj. 2	106044	81213	141196		
Inj. 3	103543	86298	142706		
Inj. 4	103884	87128	140007		
Inj. 5	98523	81752	143920		
Inj. 6	98261	89543	143904		
Average	101619	85195	142196		
Std. Dev.	3284.37	3211.56	158.85		
RSD	3.23	3.77	1.11		

## Table 3: System Precision of genotoxic impurities for Pentoxifylline

 Table 4: Method Precision of genotoxic impurities for Pentoxifylline

	% Recovery				
Sample Injection	2 shlars 1 monorel	1-bromo-3-	6-chloro-2-		
	3-chloro-1-propanor	chloropropane	hexanone		
100% recovery -1	98.10	104.90	105.10		
100% recovery-2	100.40	102.40	103.00		
100% recovery -3	102.30	102.20	99.00		
100% recovery -4	106.50	98.60	103.60		
100% recovery -5	104.20	100.70	106.60		
100% recovery -6	103.80	105.00	102.70		
Average	100.27	103.17	102.37		
RSD	2.10	1.46	3.03		

**PRECISION:** The repeatability performed the precision under the same operating conditions over a short interval of time. It expresses the closeness of agreement between a series of measurements obtained from multiple sampling from the same homogeneous sample<sup>[36]</sup>.

*a) System precision:* The system precision was determined by six replicate injections of a standard solution at 100.0% of the specified limit concerning the working concentration. Results of peak area for 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone for six replicate injections is summarized. The percentage relative standard deviation for the peak areas of 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone obtained in the range of 3.23%, 3.77%, and 1.11% at working concentration respectively.

**b)** *Method precision:* The precision of the method was determined by analyzing a sample of Pentoxifylline with 3-chloro-1-propanol, 1-bromo-3-chloropropane and 6-chloro-2-hexanone at 100% of the specification limit (six replicate spiked sample preparations). The results obtained are summarized. The percentage relative standard deviation for 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone at 100% of the specification level in six preparations should not be more than 5.0%. The percentage relative standard deviation for 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone are obtained at 2.10%, 1.46%, and 3.03 % at the working concentration. The Mean Recovery is within limits. Therefore, the

GC method for the determination of 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone in Pentoxifylline is accurate.

*LINEARITY:* Linearity solutions for the Pentoxifylline genotoxic impurities were prepared individually by diluting the stock solution at six concentration levels in the range of LOQ to 150% of the specification levels *viz.* DL, 0.3 ppm to 1.5 ppm. Tests were carried out and the slope, Y-intercept, and correlation coefficient of the calibration curve were calculated and tabulated. The peak area versus concentration was plotted. The results was linear within the range of 0.3 ppm to 1.5 ppm.









Fig. 8: Linearity plot for 6-chloro-2-hexanone



	Response Area				
Sample Name	3-chloro-1-propanol	1-bromo-3-	6-chloro-2-		
		chloropropane	hexanone		
Linearity -50%	49555	53192	72089		
Linearity - 80%	87365	76173	117673		
Linearity -100%	104754	90445	148279		
Linearity -120 %	128952	108466	172111		
Linearity -150 %	169804	130406	215030		
<b>Correlation coefficient</b>	0.995	0.994	0.995		

Table 5: Linearity summary for Pentoxifylline genotoxic impurities

**RECOVERY** (Accuracy) : Pentoxifylline was spiked with 3-chloro-1-propanol, 1-bromo-3chloropropane, and 6-chloro-2-hexanone at three different levels of 50%, 100%, and 150% of the specifications in triplicate (in total twelve determinations) and preceded according to the Sample Preparation described in Methodology ( table-6-8). Mean recovery should be in the range of 90.0% to 110.0% for 50%, 100% and 150% levels. The Mean Recovery for all components is within limits. Therefore, the GC method for the determination of 3-chloro-1propanol,1-bromo-3-chloropropane, and 6-chloro-2-hexanone in Pentoxifylline is accurate<sup>[37]</sup>.

Sample No.	Amount added (mg)	Amount recovered (mg)	% Recovery
Accuracy 50% -1	0.04900	0.05163	105.40
Accuracy 50% -2	0.04900	0.04918	100.40
Accuracy 50% -3	0.04900	0.04776	97.50
Accuracy 100% -1	0.09800	0.09660	98.60
Accuracy 100% -2	0.09800	0.10094	103.00
Accuracy 100% -3	0.09800	0.09767	99.70
Accuracy 150% -1	0.14700	0.15319	104.20
Accuracy 150% -2	0.14700	0.15094	102.70
Accuracy 150% -3	0.14700	0.15849	107.80
	Mean	102.14	
	SD	3.373	
	% RSD	3.302	

 Table 6: Recovery of 3-chloro-1-propanol

## Table 7: Recovery of f 1-bromo-3-chloropropane

Sample No.	Amount added (mg)	Amount recovered (mg)	% Recovery
Accuracy 50% -1	0.04950	0.04911	99.20
Accuracy 50% -2	0.04950	0.04930	99.60
Accuracy 50% -3	0.04950	0.05031	101.60
Accuracy 100% -1	0.09900	0.09595	96.90
Accuracy 100% -2	0.09900	0.09924	100.20
Accuracy 100% -3	0.09900	0.10172	102.70
Accuracy 150% -1	0.14850	0.14678	98.80
Accuracy 150% -2	0.14850	0.14456	97.30
Accuracy 150% -3	0.14850	0.15115	101.80
	Mean	99.79	
	SD	1.996	
	% RSD	2.000	

Sample No.	Amount added (mg)	Amount recovered (mg)	% Recovery
Accuracy 50% -1	0.04850	0.05105	105.30
Accuracy 50% -2	0.04850	0.04663	96.10
Accuracy 50% -3	0.04850	0.05024	103.60
Accuracy 100% -1	0.09700	0.10043	103.50
Accuracy 100% -2	0.09700	0.09670	99.70
Accuracy 100% -3	0.09700	0.10024	103.30
Accuracy 150% -1	0.14550	0.15051	103.40
Accuracy 150% -2	0.14550	0.15165	104.20
Accuracy 150% -3	0.14550	0.15574	107.00
	Mean	102.90	
	SD	3.202	
	% RSD	3.111	

*RUGGEDNESS* Six Pentoxifylline sample preparations of the same lot of Pentoxifylline API are made and spiked with 3-chloro-1-propanol, 1-bromo-3-chloropropane and 6-chloro-2-hexanone at 100% levels by a different analyst, using different column on a different day and injected into a different GCHS using the method as described methodology, along with Standard preparation<sup>[38]</sup>.

*a) System Precision (Ruggedness):* Six replicate injections of Pentoxifylline standard preparation will be made into the GC using the method as described under methodology.

Sr. No.	3-chloro-1-propanol		1-bromo-3-chloropropane		6-chloro-2-hexanone	
	RT	Area	RT	Area	RT	Area
1	2.70	105352	3.06	91450	4.34	154419
2	2.70	115964	3.06	92383	4.34	152895
3	2.70	111939	3.06	90118	4.34	151840
4	2.70	111810	3.06	92788	4.34	165708
5	2.70	114398	3.06	91124	4.34	155678
6	2.70	117426	3.06	91136	4.34	155946
Average	2.70	112815	3.06	91500	4.34	156081
Std. Dev.		4269.83		961.76		4975.11
RSD		3.78		1.05		3.19

 Table 9: Ruggedness - System precision

 Table 10: Ruggedness - 100% Spike of standard solution

	% Recovery			
Sample Injection	3-chloro-1-propanol	1-bromo-3- chloropropane	6-chloro-2-hexanone	
100% recovery -1	105.40	101.20	104.80	
100% recovery-2	102.30	101.90	105.30	
100% recovery -3	100.50	103.40	103.40	
100% recovery -4	102.10	103.80	107.80	
100% recovery -5	102.80	100.40	107.80	
100% recovery -6	105.50	103.70	103.90	
Average	102.73	102.17	104.50	
RSD	2.41	1.10	0.94	

		% Recovery				
Analyst No	Sample Injection	3-chloro-1-	1-bromo-3-	6-chloro-2-		
		propanol	chloropropane	hexanone		
	100% recovery -1	98.10	104.90	105.10		
	100% recovery-2	100.40	102.40	103.00		
	100% recovery -3	102.30	102.20	99.00		
Analyst I	100% recovery -4	106.50	98.60	103.60		
Analyst I	100% recovery -5	104.20	100.70	106.60		
	100% recovery -6	103.80	105.00	102.70		
	100% recovery -1	105.40	101.20	104.80		
	100% recovery-2	102.30	101.90	105.30		
	100% recovery -3	100.50	103.40	103.40		
Analyst II	100% recovery -4	102.10	103.80	107.80		
	100% recovery -5	102.80	100.40	107.80		
	100% recovery -6	105.50	103.70	103.90		
	<b>Overall</b> Average	102.83	102.35	104.42		
	<b>Overall Std. Dev.</b>	2.4275	1.9233	2.4368		
	<b>Overall RSD</b>	2.36	1.88	2.33		

Table 11: Ruggedness - 100% Spike of standard solution (Analyst I &II)

The RSD of system precision (ruggedness) 3-chloro-1-propanol is 3.78%, 1-bromo-3chloropropane is 1.05 % and 6-chloro-2-hexanone is 3.19 % respectively and it meets acceptance criteria. Therefore, the GC method for the determination of 3-chloro-1-propanol, 1bromo-3-chloropropane and 6-chloro-2-hexanone in Pentoxifylline API is precise.

**b)** Method Precision (Ruggedness) : Five sample preparations of Pentoxifylline are to be prepared as the procedure given in the method and injected into the GC using the method as described under Methodology. Based on above study shows that the reported impurities of 3-chloro-1-propanol, 1-bromo-3-chloropropane, and 6-chloro-2-hexanone in Pentoxifylline API is reproducible.

c) Intermediate Precision (Spike Study): Spike 3-chloro-1-propanol, 1-bromo-3chloropropane, and 6-chloro-2-hexanone at a 100% concentration of specification level in Pentoxifylline substance and inject and process as per the methodology (Table-10-11). Mean recovery should be in the range of 90.0% to 110.0% for 100% levels. The Mean Recovery is within limits. Therefore, the GC method for the determination of 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol in Pentoxifylline is suitable to use in routine analysis. % recovery of all impurities for both Analyst are well with the defined specification for 6chloro-2-hexanone is 2.33%, 1-bromo-3-chloropropane is 1.88% and 3-chloro-1-propanol is 2.36% hence the GC method for the determination of 6-chloro-2-hexanone, 1-bromo-3chloropropane and 3-chloro-1-propanol in Pentoxifylline API is reproducible. **ROBUSTNESS:** Prepared samples at a 100% concentration of specification level in the sample and injected and processed as per the methodology. Separately standard preparations containing a concentration of 100% specification level of 6-chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol are also to be prepared and injected and used for quantification of impurities in the sample a) As such method, b) Change in flow rate ( $\pm 2\%$ )

		% Recovery		
Analyst No	Sample Injection	3-chloro-1-	1-bromo-3-	6-chloro-2-
		propanol	chloropropane	hexanone
	100% recovery -1	98.10	104.90	105.10
	100% recovery-2	100.40	102.40	103.00
	100% recovery -3	102.30	102.20	99.00
Analyst –I	100% recovery -4	106.50	98.60	103.60
(as such	100% recovery -5	104.20	100.70	106.60
method)I	100% recovery -6	103.80	105.00	102.70
	100% recovery -1	92.80	95.00	102.50
	100% recovery-2	98.00	97.40	97.50
	100% recovery -3	106.50	98.50	96.60
Analyst –II	100% recovery -4	102.30	97.90	100.20
(change in	100% recovery -5	104.90	92.50	101.60
flow)	100% recovery -6	101.90	99.30	100.70
	<b>Overall Average</b>	101.80	99.53	101.59
	<b>Overall Std. Dev.</b>	3.9903	3.7490	2.9629
	<b>Overall RSD</b>	3.92	3.77	2.92

 Table 12: Robustness - 100% Spike of standard solution (With different Analyst and change in condition)

**CONCLUSION:** Three genotoxic impurities may be present in the synthesis of Pentoxifylline. The proposed method was found to be specific, accurate, linear, precise, and very useful for monitoring three genotoxic impurities -chloro-2-hexanone, 1-bromo-3-chloropropane, and 3chloro-1-propanol in pharmaceutical analysis. The % recovery of all impurities for both Analyst with different chromatographic conditions are well with the defined specification for 6-chloro-2hexanone is 2.92%, 1-bromo-3-chloropropane is 3.77% and 3-chloro-1-propanol is 3.92% hence the GC method for the determination of 6-chloro-2-hexanone, 1-bromo-3-chloropropane and 3chloro-1-propanol in Pentoxifylline API is robust. The purpose of this study is to quantify and qualify suitable methods and threshold levels of the three genotoxic impurities that may be present in the synthesis of Pentoxifylline. A gas chromatographic technique with an FID detector was adopted for suitable quantification of these impurities. The proposed method was found to be specific, accurate, linear, precise, and very useful for monitoring three genotoxic impurities -chloro-2-hexanone, 1-bromo-3-chloropropane, and 3-chloro-1-propanol in pharmaceutical analysis. Genotoxic alerts on structures of these impurities were confirmed by the Genotoxic Predication report.

**ACKNOWLEDGMENT:** The authors are thankful to Sathaye College, Mumbai for Providing a research laboratory, and for providing the gift sample for this Research work by Supriya Life Sciences, Mumbai. The authors want to acknowledge the contributions of the Department of Chemistry, Pacific University Udaipur.

**COMPETING DECLARATION:** The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this research paper.

#### REFERENCES

- Atturu G, Homer-Vanniasinkam S, Russell DA. Pharmacology in Peripheral Arterial Disease: What the Interventional Radiologist Needs to Know. *Seminar in Interventional Radiology*. 2014; 31: 330–337. doi: 10.1055/s-0034-1393969.
- **2.** Generic Drug Facts, Center for Drug Evaluation and Research, U.S. Food and Drug Administration, (2017).
- Hansen K, Mika S, Schroeter T, Sutter A, Laak A, Steger-Hartmann T,et.al. Benchmark data set for in Silico prediction of Ames mutagenicity. *Journal of Chemical Information and Modeling*. 2009; 49: 2077-2081. doi:: 10.1021/ci900161g.
- 4. Sasaki JF, Sekihashi K, Izumiyama F, Nishidate E, Saga A, Ishida K. The Comet assay with multiple mouse organs: comparison of Comet assay results and carcinogenicity with 208 chemicals selected from the IARC monographs and U.S. NTP carcinogenicity database. *Critical Reviews in Toxicology*. 2000; 30: 629-799. doi:10.1080/10408440008951123.
- Monteiro JP, Alves MG, Oliveira PF, Silva BM. Structure-Bioactivity Relationships of Methylxanthines, Trying to Make Sense of All the Promises and the Drawbacks. *Molecules*.2016: 21(974): 1-32. doi:10.3390/molecules21080974.
- Yan K. Li-Na G, Yuan-Lu C, Zhang Y, Zhou X. The cyclic AMP signalling pathway: Exploring targets for successful drug discovery (Review). *Molecular Medicine Reports*. 2016;13: 3715–3723.doi: 10.3892/mmr.2016.5005
- Churi SK, Lokhande MV. Impurity Profiling of Pharmaceutical Drugs by Various Methods. Journal of Applied Chemistry. 2017; 10(7):27-34.
- **8.** Madan RK, Levitt J. A review of toxicity from topical salicylic acid preparations. *Journal of the American Academy of Dermatology*.2014; 70 : 788-792. doi: 10.1016/j.jaad.2013.12.005
- **9.** Gupta MK, Lokhande MV, Structure Determination of Impurity In Memantine Hydrochloride By Analytical Techniques. *International Journal of General Medicine and Pharmacy.* 2014;3(4): 75-84.
- **10.** Goldberg DR. Aspirin: Turn of the Century Miracle Drug. *Chemistry Heritage Magazine*. 2009: 27:26-30.

- 11. Van Wagner LB, Koppe WP, Brunt EM, Gottstein J, Gardikiotes K, Rinella ME. Pentoxifylline for the treatment of non-alcoholic steatohepatitis: a randomized controlled trial. *Annals of Hepatology*. 2011;10:277-286.doi: 10.1016/S1665-2681 (19) 31539-X
- Zate PB, Kothari S, Lokhande MV. Confirmation and Quantification of Genotoxic Impurity 2-Dimethyl-aminoethyl chloride hydrochloride by GCMS in Chlorpheniramine/ Chlorphenamine Maleate. *Journal of Applied Chemistry*. 2017; 10(7): 21-26.
- 13. Hoffman R. Goldfrank's Toxicologic Emergencies, 10<sup>th</sup> edn 2015: 915.
- Belsito D, Bickers D, Bruze M. Toxicologic and Dermatologic Assessments for Three Groups of Fragrance Ingredients, *Food Chemical Toxicology*. 2007:45(1):S130-67. doi: 10.1016/j.fct.2007.09.067 (2007) 45.
- 15. Aggarwal K, Gautam M, Singh M, Kharat N, Singh V, Vyas S, Singh HP. Prophylactic Use of Pentoxifylline and Tocopherol in Patients Undergoing Dental Extractions Following Radiotherapy for Head and Neck Cancer. *Nigerian Journal of Surgery* .2017; 23:130–133. doi: 10.4103/njs.NJS\_40\_16.
- Kazius J, McGuire R, Bursi R. Derivation and Validation of Toxicophores for Mutagenicity Prediction. *Journal of Medicinal Chemistry*. 2005;48:312-320. doi:10.1021/jm040835a.
- **17.** Rathod NG, Lokhande MV. Characterisation and Identification of Process-Related Impurity in Amodiaquine. HCl by Using Some Analytical Techniques: A Review, *American Journal of Advanced Drug Delivery*. 2015; 3(5): 2015.
- Maurya CP, Lokhande MV. Characterization and validation of impurities related to the pharmaceutical bulk drug (API) by using some analytical techniques. Inter J Pharm Sci Res. 2017;8(8):3325-3340. doi: 10.13040/IJPSR.0975-8232.8(8).3325-40.
- Williams K, Sobol RW. Mutation Research / Fundamental and Molecular Mechanisms of Mutagenesis Special Issue: DNA Repair and Genetic Instability. *Mutation Research*.2013: 1-3: 743-744. doi:10.1016/j.mrfmmm.2013.04.009.
- 20. Paul N, Kennedy A, Franco N, Hezekiah K. A Versatile HPLC Method for the Simultaneous Determination of Bromhexine, Guaifenesin, Ambroxol, Salbutamol, Terbutaline, Pseudoephedrine, Triprolidine, and Chlorpheniramine Maleate in cough-cold syrups. Chromatographia.2016;79(21):1507-1514.<u>doi: 10.1007/s10337-016-3158-1.</u>
- Ramakrishna K, Praveen B. Validated Stability Indicating RP-HPLC Method for Simultaneous Estimation of Codeine Phosphate and Chlorpheniramine Maleate from Their Combined Liquid Dosage Form. *Chromatography Research International*. 2013:2013:1-7.doi:<u>10.1155/2013/404727</u>.

- **22.** Larisa A, Midhat V, Edina C, Mirsad D. Development and Validation of an HPLC Method for Chlorphenamine Maleate Related Substances in Multicomponent Syrups and Tablets. *International Journal of Pharmacy Teaching and Practices*. 2014;5: 997-1001.
- **23.** Hanan S, Amir AS, Saleh T. Separation and Assay of Four Antihistamine drugs Diphenhydramine, Chlorpheniramine, Cyproheptadine and Fexofenadine in Pharmaceutical forms by a Single HPLC Method. *International Journal of Pharmacy & Pharmaceutical Sciences*. 2018;10: 53-60. doi:10.22159/ijpps.2018v10i4.24819.
- 24. Ambadekar SR, Balakrishnan I, Lokhande MV. Validation of Pharmaceutical (API) Bulk Drug by HPLC Methods. *Journal of Applied Chemistry*. 2018; 11(2): 01-20. doi:10.9790/5736-1102020120
- 25. Lakhmapure SB, Kothari S, Lokhande MV. Validation of gas chromatography (GC) method for residual solvent in brompheniramine maleate (API). Inter J Pharm Sci Res. 2020; 11(10) :(2020),pp.5039-5052.doi:10.13040/IJPSR.0975-8232.11(10). 5039-52.0
- 26. Churi SK, Lokhande MV, Identification and Impurity Profiling of Process Related Impurities In DTPEE, *European Journal of Biomedical and Pharmaceutical Sciences*. 2017; 4(9): 617-623.
- 27. Wagh SS, Kothari S, Lokhande MV. Quantification Of (4-Bromophenyl) {Pyridine-2-Yl}Acetonitrile Impurity (4-BPPA) by HPLC In Bromopheniramine Maleate Active Pharmaceutical Ingredient. *Journal of Applied Chemistry* .2017;10(10):26-31. doi:10.9790/5736-1006022 631.
- **28.** Zate PB, Kothari S, Lokhande MV. Determination and Quantification of Carryover Genotoxic Impurities 2-Chloropyridine (2CP) and 4-Bromobenzyl Cyanide by GCHS in Brompheniramine Maleate API. *International Journal of Pharmacy and Pharmaceutical Research*. 2017;10(7): 13-24.
- **29.** Hanan S, Amir AS, Saleh T. Separation and Assay of Four Antihistamine drugs Diphenhydramine, Chlorpheniramine, Cyproheptadine and Fexofenadine in Pharmaceutical forms by a Single HPLC Method. *International Journal of Pharmacy and Pharmaceutical Science*. 2018;10: 53-60. <u>doi:10.22159/ijpps.2018v10i4.24819</u>
- **30.** Manassra A, Khamis M, El-Dakiky M, Abdel-Qader Z, Al-Rimawi F. Simultaneous HPLC analysis of pseudoephedrine hydrochloride, codeine phosphate, and triprolidine hydrochloride in liquid dosage forms. *Journal of Pharmaceutical and Biomedical Analysis*. 2011; 51: 991-993. doi: 10.1016/j.jpba.2009.10.024
- **31.** https://www.drugs.com/international/pentoxifylline.
- **32.** ICH Q3A and Q3B (R2), Impurities in New Drug Substances. 2006.

- **33.** ICH M7, Assessment and Control of DNA Reactive (Mutagenic) Impurities in Pharmaceuticals to Limit Potential Carcinogenic Risk, Business Plan.2010.
- 34. Li W, Zheng L, Sheng C, Cheng X, Qing L, Qu S. Systematic review on the treatment of pentoxifylline in patients with non-alcoholic fatty liver disease. *Lipids Health Diseases*. 2011; 10: 1-67. doi: 10.1186/1476-511X-10-49.
- **35.** Lokhande MV, Rathod NG, Gupta MK. Structural elucidation, Identification, and quantization of process-related impurity in Hydralazine Hydrochloride HR/AM- LC-MS/MS, NMR, and FTIR technique. *Journal of Applied Chemistry*. 2013;6(2):05-15.
- 36. Gorajiya A, Shelat P, Lalwani A. Formulation and Characterization of Imatinib Mesylate Liposomes in Gel for Intraarticular Administration. International Journal of Pharmaceutical Science and Drug Research. 2022;14(5):559-566. doi:10.25004/ IJPSDR.2022.140508
- Lakhmapure SB, Telavane SA, Kothari S, Lokhande MV. Method of Validation for Residual Solvents in Brimonidine tartrate by GC-HS. J Appl Chem. 2023; 12 (1): 42-52.

\*\*\*\*\*